

# **Doing Thousands of Hypothesis Tests at the Same Time**

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# Simultaneous Hypothesis Testing

**1980:** “Simultaneous Statistical Inference” (Rupert Miller)

- 2, 3, ..., 20 simultaneous tests

**Today:** Several thousand tests

- *High Throughput Devices:*  
Microarrays, fMRI, proteomics, large-scale surveys
- **Love/Hate** Classical single-test theory
- frequentist, Bayes, empirical Bayes

## A Microarray Example: The Prostate Data (Singh et al. 2002)

- *102 Subjects*: 50 normal, 52 cancer
- $N = 6033$  genes :  $X$   $6033 \times 102$
- *Which genes are “non-null”?*  
i.e. expressed differently in cancer vs normal subjects?

## ***t*-statistics and *z*-scores**

- $i^{\text{th}}$  row of  $X$   
normals  $(x_{i1}, x_{i2}, \dots, x_{i50})$   
cancer  $(x_{i51}, x_{i52}, \dots, x_{i102})$  } “ $t_i$ ”
- $t_i$  = two-sample  $t$ -stat, cancer vs normals

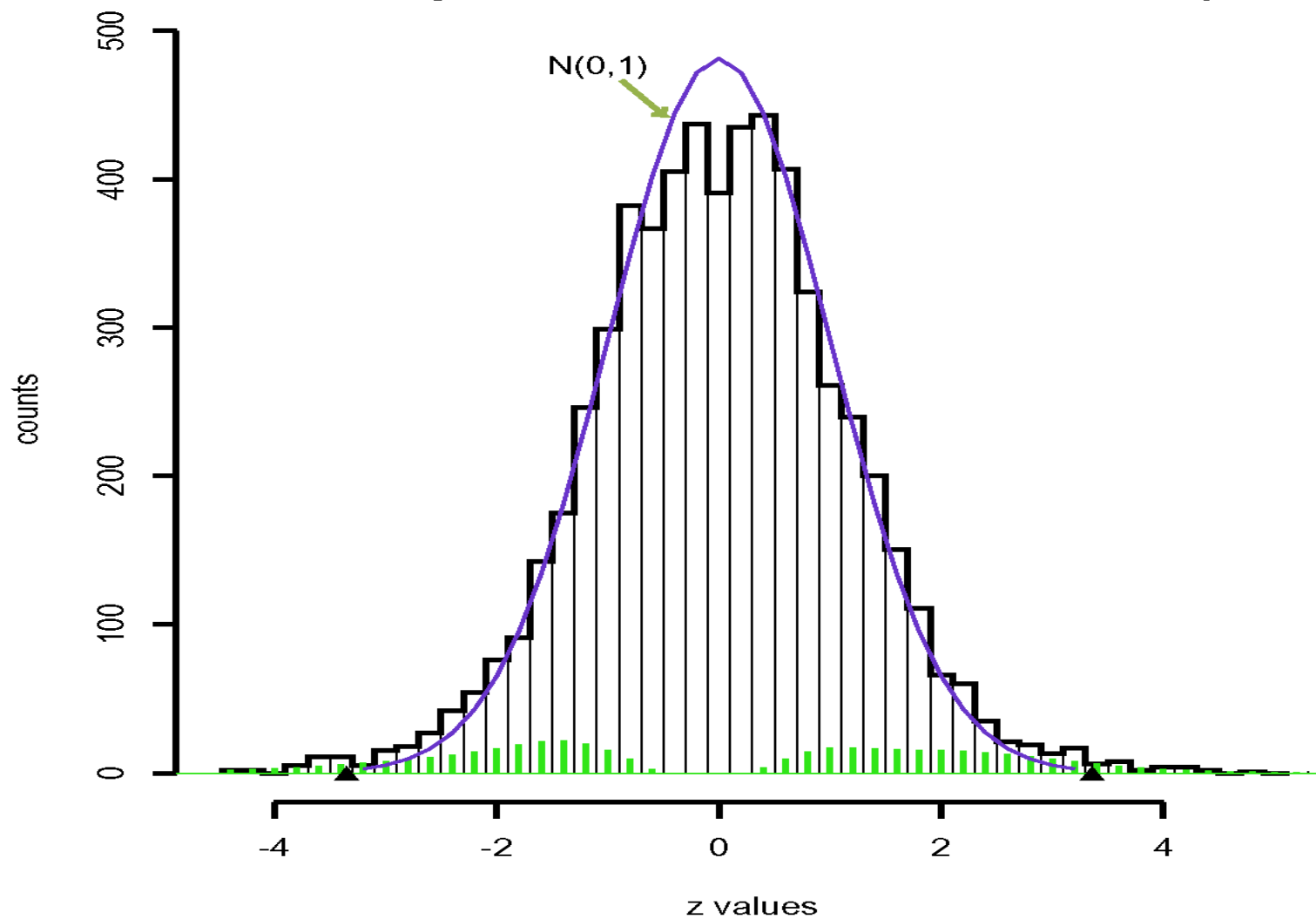
$z$ -scores

$$z_i = \Phi^{-1}(F_{100}(t_i))$$

where  $\phi$  is  $N(0, 1)$  cdf, and  $F_{100}$  is cdf for  $t_{100}$ .

- **Theoretical Null**       $z_i \sim N(0, 1)$

Prostate Data, Singh et al: z-values for 6033 genes,  
Comparing 50 normals with 52 cancer subjects



## The Two-Groups Model

*Two Classes of Genes* “null”, “non-null”

- $\begin{cases} p_0 = \text{Prob \{null\}}, & f_0(z) \text{ density if null} \\ p_1 = \text{Prob \{non-null\}}, & f_1(z) \text{ density if non-null} \end{cases}$
- $p_0$  large ( $\geq 0.90$ )
- **Theoretical Null**  $f_0 \sim N(0, 1)$

(fits center of histogram)

- $f_1(z)$  ”long tailed” ( $z_i$ ’s far from zero)

## False Discovery Rates (Efron 2006)

- *Mixture density*

$$f(z) = p_0 f_0(z) + p_1 f_1(z)$$

### Bayes Rule “Local false discovery rate”

$$\begin{aligned} fdr(z) &= Prob\{gene\ i\ null | z_i = z\} \\ &= p_0 f_0(z) / f(z) \end{aligned}$$

$Fdr(z)$  Replace densities with cdfs

# Empirical Bayes

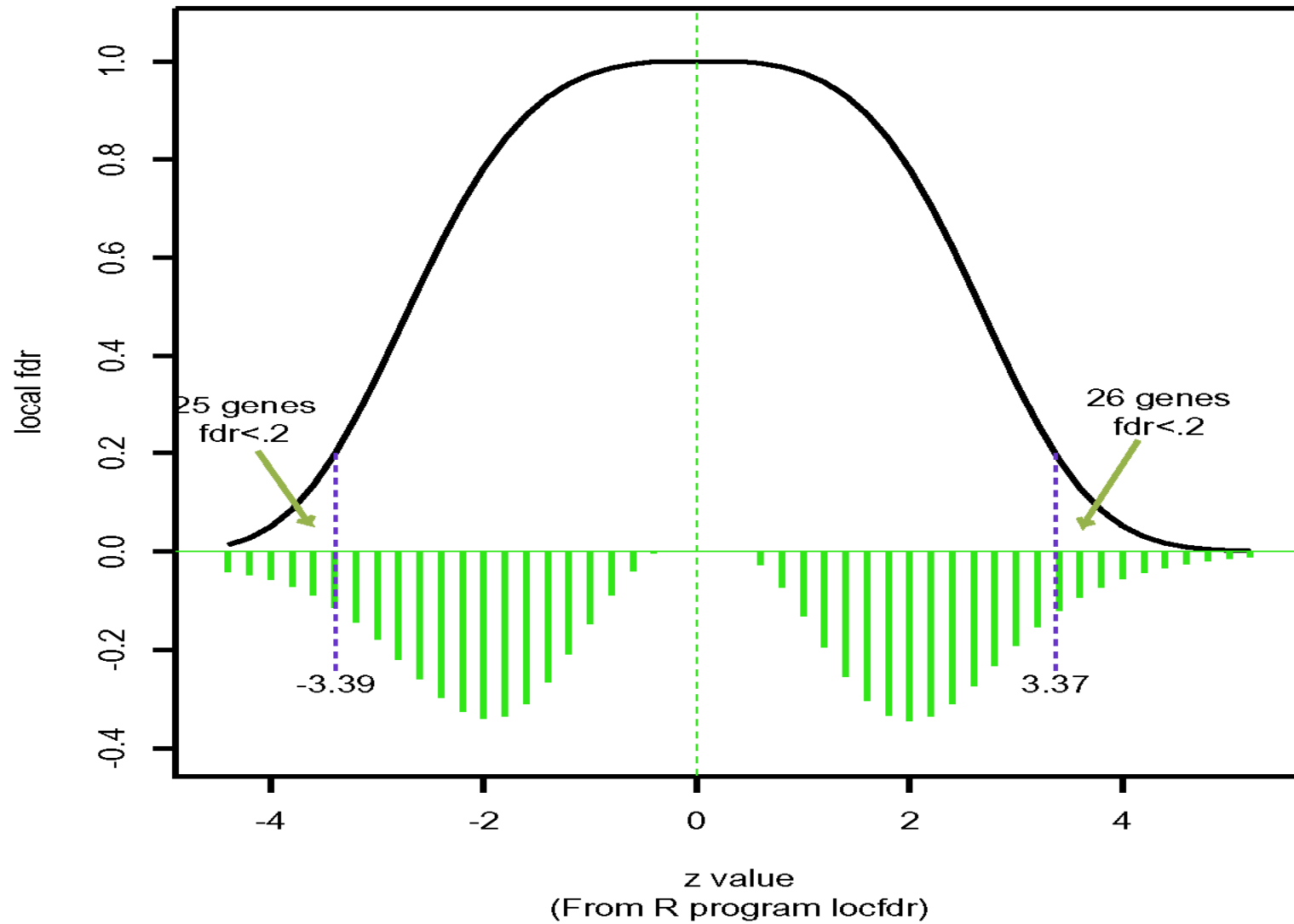
- Estimate mixture density  $f(z)$  from observed z-values  $z_1, z_2, \dots, z_N$  :

$$\widehat{fdr}(z) = p_0 f_0(z) / \widehat{f}(z)$$

- *BH* :  $p_0 = 1$  ( $\widehat{p}_0 = 0.94$ .)
- *Don't need*:  $z_i$ 's independent, t-tests ...



Estimated  $fdr(z)$  for prostate data (solid curve)  
Bars proportional to non-null histogram



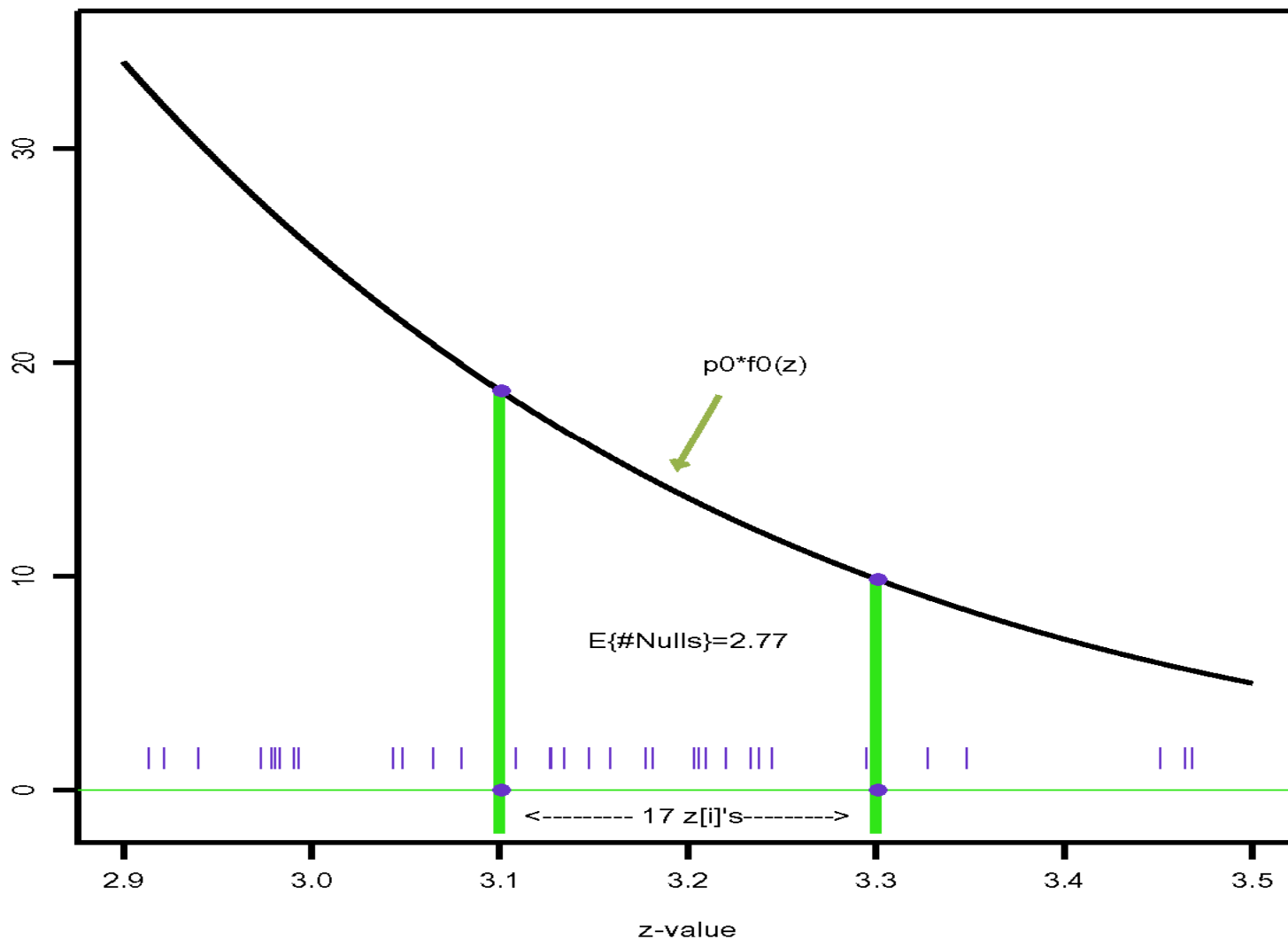
## Basic Fdr Idea

- Histogram has 49 bins, width  $\Delta = .2$
- $\text{bin}_{39} = [3.1, 3.3]$ ; count  $y_{39} = 17$
- $y_{39}^{(o)} = \# \{\text{null genes in bin}_{39}\} = ??$
- $\hat{y}_{39}^{(o)} = E\# \{\text{null genes in bin}_{39}\} = 2.77$   
(=  $6033 \cdot 0.94 \cdot \varphi(3.2) \cdot 0.1$ )

- $\widehat{fdr}(3.2) = 2.77/17 = .16$

- *About one sixth of the 17 genes in  $\text{bin}_{39}$  are false discoveries*

close-up of histogram:  $fdr(3.2) = 2.77/17$



## The Non-Null Counts

- $y_k$  = observed count in  $k^{th}$  bin
- $1 - fdr(z_i) = Prob\{gene\ i\ non\ null\ |z_i = z\}$
- So estimated number non-nulls in  $bin_k$  is

$$\hat{y}_k^{(1)} = [1 - \widehat{fdr}_k] y_k$$

where  $\widehat{fdr}_k = \widehat{fdr}(z = \text{midpoint } bin_k)$ .

- Plotted bars used smoothed version.

## Power Diagnostics (Efron 2006, Section 3)

Good Power:  $\widehat{fdr}_k$  small where  $\widehat{y}_k^{(1)}$  big

Expected Non-Null  $fdr$

$$E\widehat{fdr}^{(1)} = \sum \widehat{fdr}_k \cdot \widehat{y}_k^{(1)} / \sum \widehat{y}_k^{(1)}$$

Prostate Data  $E\widehat{fdr}^{(1)} = 0.68$  (Bad!)

- “Why aren’t our favorite genes on your list of non-null cases?”

## Increased Sample Size

- Multiply number microarrays by “ $c$ ”

( $c = 2 : 100$  non-tumor men, 104 tumor)

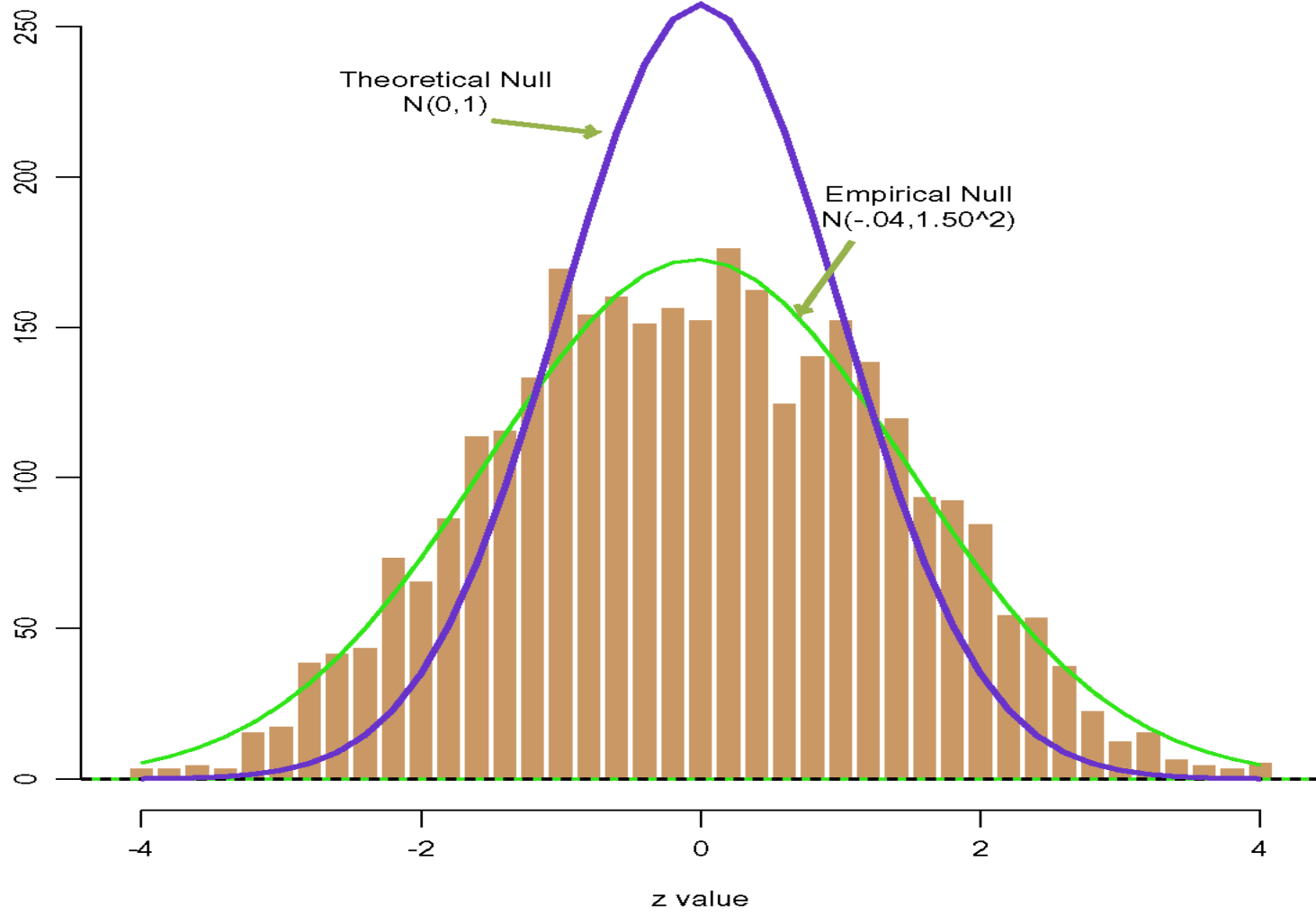
- Can estimate improvement in  $\widehat{E}fdr^{(1)}$  :

$c$ :	1	1.5	2	2.5	3
$\widehat{E}fdr^{(1)}$	.68	.54	.44	.38	.34

## The BRCA Data (Hedenfalk et al. 2001)

- *Microarray study* comparing tumors from women with BRCA1 or BRCA2 mutations
- *15 microarrays*: 7 BRCA1, 8 BRCA2,  
same 3226 genes:  $X$   $3226 \times 15$
- $t_i =$  two-sample  $t$  stat for gene  $i$ , BRCA2 vs BRCA1
- $z_i = \Phi^{-1}(F_{13}(t_i))$
- *Theoretical Null*  $z_i \sim N(0, 1)$

# BRCA data: 3226 z-values





## Four Arguments Against the Theoretical Null

- Central histogram doesn't match theoretical  $N(0, 1)$  null  
(requires  $p_0 \leq 0.7$ ).
- Central hist matches “empirical null”  $N(-.04, 1.50^2)$
- Four Reasons Why null  $z_i$ 's *not*  $N(0, 1)$
- **Reason I Failed Assumptions:**
- Maybe nonnormality of microarray measurements Distorts student's  $t$  distribution for “ $t_i$ ”
- *Permutation Null:* Scramble the 102 microarrays  
(gives  $z_i \sim N(0, 1)$  for prostate data)

## Reason 2: Unobserved Covariates (Efron 2004, Section 4)

- *BRCA Study Observational* Unobserved Covariates  
Age, Wt, Stage, Race ...
- *If observed* would be factored out of  $z_i$ 's
- Tends to *widen* null density  $f_0(z)$
- Could account for BRCA histogram
- *Won't show up in permutation distribution*

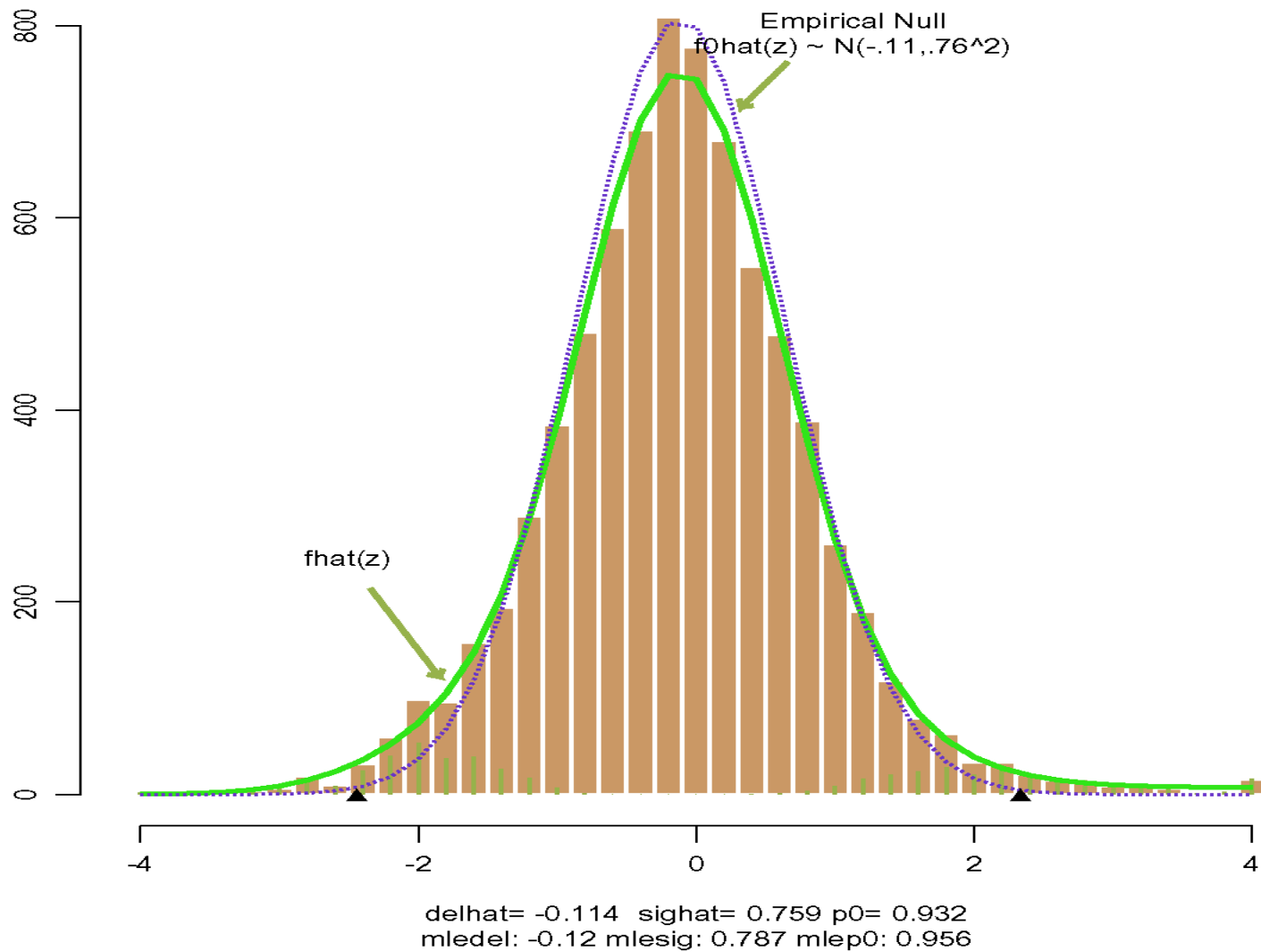
## Reason 3: Correlation Across Arrays

- *Gene i data* :  $(x_{i1}, x_{i2}, \dots, x_{i7}), (x_{i8}, x_{i9}, \dots, x_{i15})$
- Student-*t* null density assumes independence across microarrays
- Principle Component Analysis showed correlation among  $(x_{i8}, x_{i9}, x_{i10}, x_{i11})$ , likewise  $(x_{i12}, x_{i13}, x_{i14}, x_{i15})$
- “Less than 13 df”
- Not detectable from permutations

## Reason 4: Correlation Across Genes (Efron 2007)

- $\widehat{fdr}(z) = p_0 f_0(z) / \widehat{f}(z)$  does **not** require independence of gene measurements.
- *However: gene-wise correlations affect “ $f_0$ ”*
- *BRCA: 5 million pairwise correlations  
rms correlation = 0.15*
- *Even if  $f_0(z_i) \sim N(0, 1)$  individually,  
can behave as  $N(\theta, \sigma_0^2)$  collectively,  
with  $.75 < \sigma_0 \leq 1.25$ .*
- **Not detectable from permutations.**

HIV data: compare 4 HIV+ subjects with 4 HIV- subjects,  
N=7680 genes; van't Wout et al (2003)

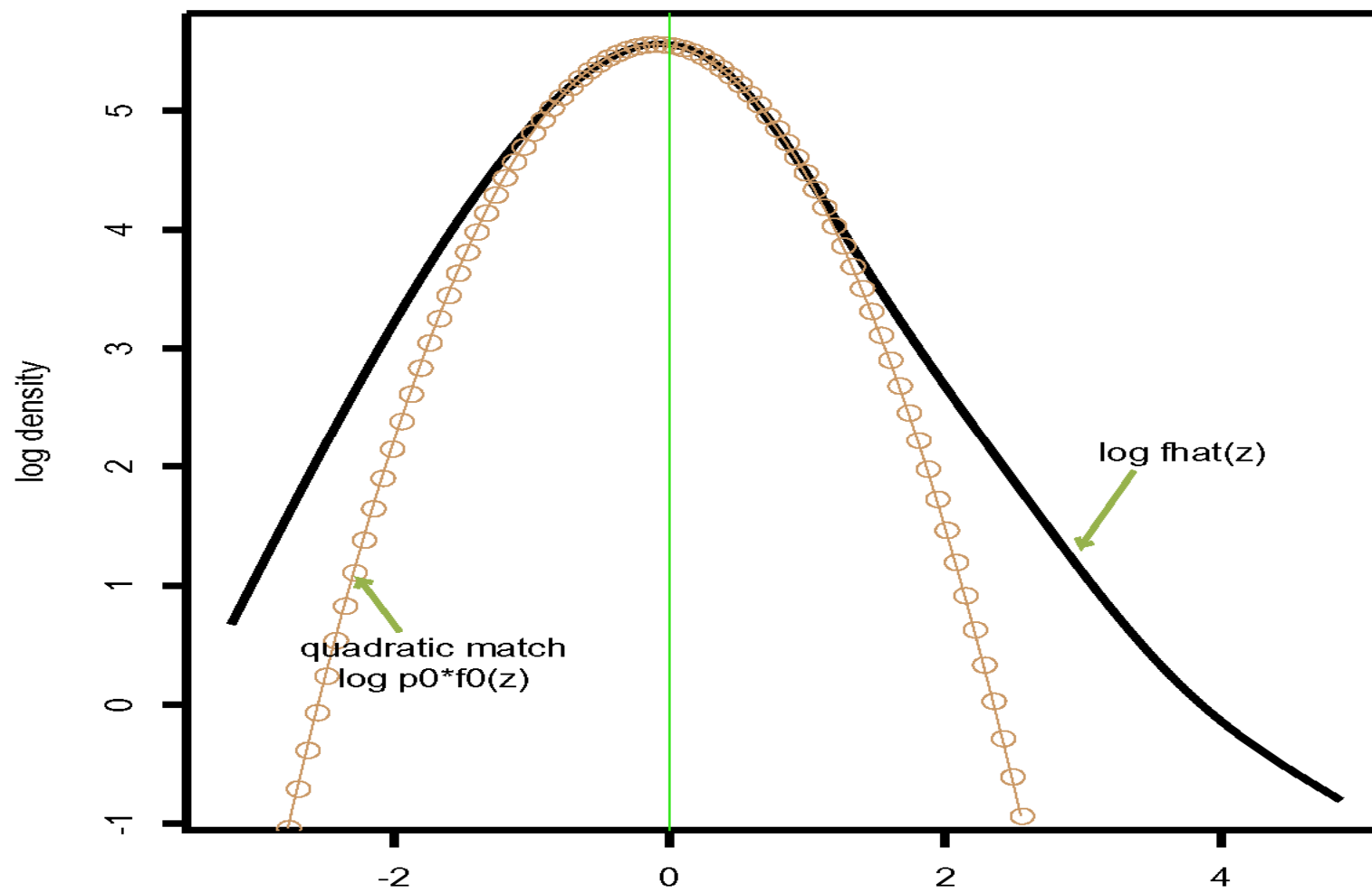


## Empirical Null Estimation

$$\text{fdr}(z) = p_0 f_0(z) / f(z)$$

- Theoretical  $N(0, 1)$  may not fit  $z$ -value histogram central peak
- *Idea* Assume  $f_0(z) \sim N(\delta_0, \sigma_0^2)$ ; fit  $p_0, \delta_0, \sigma_0$  from histogram counts near  $z = 0$ .    [“zero assumption”]
- *Central Matching* :
  - (1) Plot  $\log \hat{f}(z)$
  - (2) Find best quadratic match near  $z = 0$
  - (3) coeffs of match  $\Rightarrow (\hat{p}_0, \hat{\delta}_0, \hat{\sigma}_0)$
- Nearly unbiased for  $(\hat{\delta}_0, \hat{\sigma}_0)$ , Efron (2004).

Estimate  $(p_0, \delta_0, \sigma_0)$  by quadratic fit  
to  $\log \hat{f}(z)$  around  $z=0$



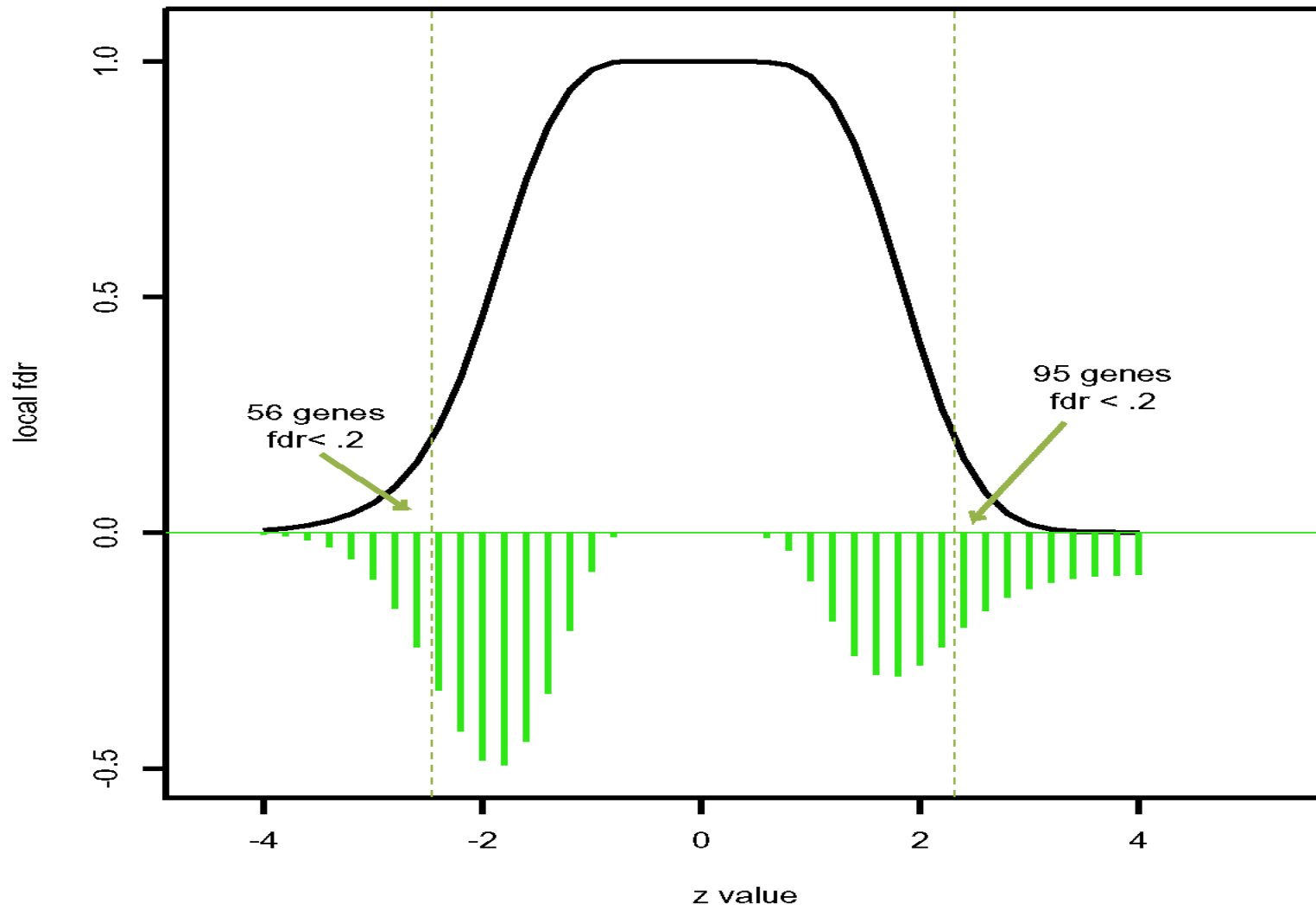
$p_0\text{hat}=.93, \delta_0\text{hat}=-.11, \sigma_0\text{hat}=.76$

## Direct Maximum Likelihood Estimation of $(p_0, \delta_0, \sigma_0)$

- Assume all the  $z_i$ 's in  $[-x_0, x_0]$  are from  $f_0(z)$ , the null density
- Let  $\mathbf{Z}_0 = (Z_1, Z_2, \dots, Z_{N_0})$  be them.
- Then  $\mathbf{Z}_0$  follows a truncated  $N(\delta_0, \sigma_0^2)$  distribution: can estimate  $(p_0, \delta_0, \sigma_0)$ .
- More biased, less variable than central matching method



Estimated  $fdr(z)$  for HIV data (solid curve)  
Bars proportional to non-null histogram ( $Efdr1=.46$ )



# Large-Scale Simultaneous Testing

- *Not just a lot of classical single tests*
  - Multiplicities
  - Empirical Bayes
- Can learn things you didn't want to know
- Permutation methods not cure-alls
- **Modelling:** Better to minimize
  - Model inside of  $X$  ?
  - Big data sets should supply own models

## References

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- Van't Wout et al. (2003). Cellular human gene expression upon human immunodeficiency versus type I infection of CDS + T-Cell lines. *J. Virol.* 1392–1402.
- “locfdr” R program, available on CRAN on Efron site above.